

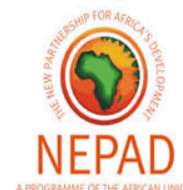
*Advanced Genomics - Bioinformatics  
Workshop*

**Mark Wamalwa**

*BecA-ILRI Hub, Nairobi, Kenya*

<http://hub.africabiosciences.org/>

[m.wamalwa@cgiar.org](mailto:m.wamalwa@cgiar.org)

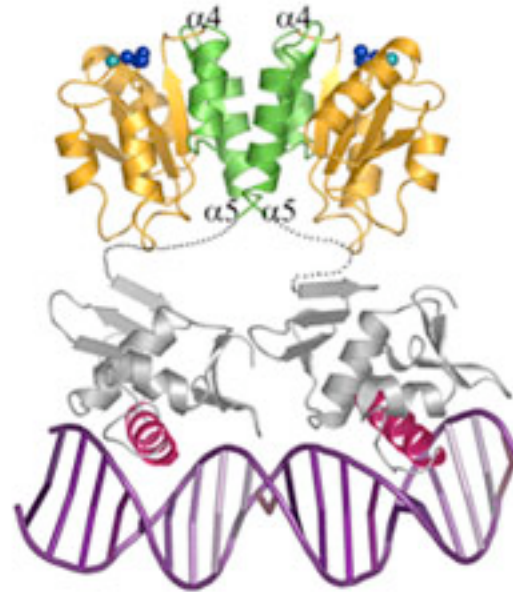


7<sup>th</sup> – 18<sup>th</sup> September 2015

**biosciences**

eastern and central **africa**

## Evolution and constraint on *cis*-regulatory motifs (focusing on TF binding sites)

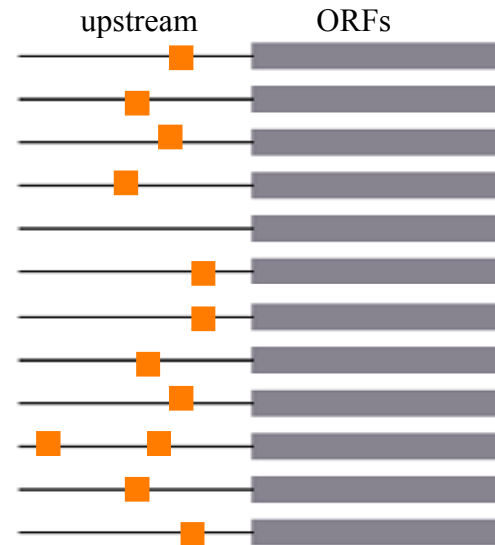
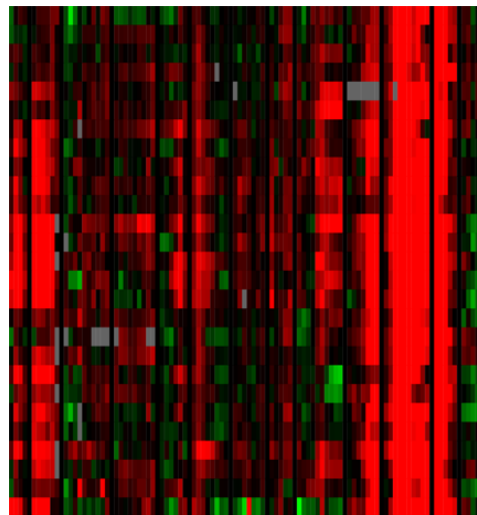


Many DNA binding proteins recognize specific (often short) DNA sequences.

Often bind 'degenerate' sequences, since some bases more important for contact.

Many work cooperatively with other factors to bind.

# Representing the set of TF binding sites *within* a genome



Site 1	A	G	A	T	G	G	A	T	G	G
Site 2	T	G	A	T	T	G	A	T	G	T
Site 3	T	G	A	T	G	G	A	T	G	G
Site 4	A	G	A	T	T	G	A	T	C	G
Site 5	T	G	A	T	G	G	A	T	T	G
Site 6	T	G	A	T	G	G	A	T	T	G
Site 7	A	G	A	T	G	G	A	T	T	G

IUPAC consensus:

**W G A T G G A T N G**

## Position-weight matrices are a better representation

Site 1	A	G	A	T	G	G	A	T	G	G
Site 2	T	G	A	T	T	G	A	T	G	T
Site 3	T	G	A	T	G	G	A	T	G	G
Site 4	A	G	A	T	T	G	A	T	C	G
Site 5	T	G	A	T	G	G	A	T	T	G
Site 6	T	G	A	T	G	G	A	T	T	G
Site 7	A	G	A	T	G	G	A	T	T	G

---

**PWM represents frequencies of each base at each position in the motif**

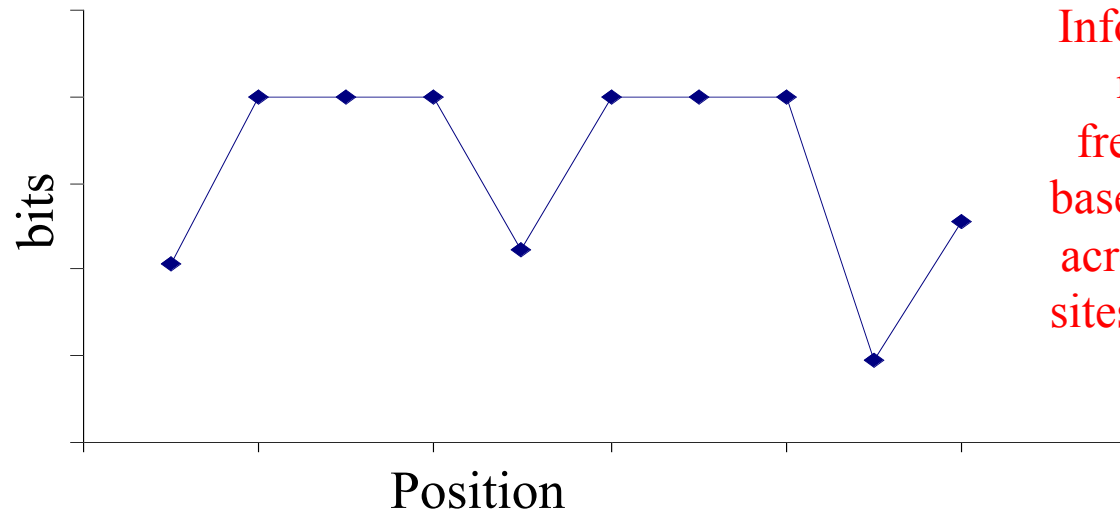
G	0	1.0	0	0	0.7	1.0	0	0	0.4	0.8
A	0.4	0	1.0	0	0	0	1.0	0	0	0
T	0.6	0	0	1.0	0.3	0	0	1.0	0.4	0.2
C	0	0	0	0	0	0	0	0	0.2	0

# Web-logo: A graphical representation of PWMs

<http://weblogo.berkeley.edu/>



Information Profile:



Information content represents the frequency of each base at each position across ALL binding sites in an individual

To study the evolution of cis regulatory elements, we first need to identify them in genomes

### Identification of *cis*-regulatory elements

Computational predictions:

1. Scan genome for matches to known matrix/consensus  
*problem is that there are many nonfunctional in the genome - poor predictor of function*
2. Phylogenetic footprinting: overly-conserved sequences in multiple alignments  
*Variation within element is typically lower than surrounding 'nonfunctional' DNA*

Simplest case: stretches of very highly conserved sequence

**TATA**

```

CTAATCTTAC TTATA TGTGGT-GGAAAT-GTAAAGAGCCCCATTATCTTAGCCTAAAAAACC--TTCTCTTTGGAACTTTCAG!
CTAGTCTTAC TTATA TGTGGT-GAGAGT-GTTGATAACCCAGTATCTTAACCCAAGAAAGCC--TT-TCTATGAAACTTGAAC!
CTAGTCTTAC TTATA TGTGGT-GGGAATTGTTGGTAATCCAGTCTCCCAGATCAAAAAAGGT--CTTTCTATGGAGCTTTG-C!
CTGATCTTCT TTATA TATTATAGAGAGATGCCAATAAACGTGTACTACCTCGAACAAAAGAAGGGGATTTTCTGTAGGGCTTTCCC!
**  ****  *****  **  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *

```

**Gal4** **Gal4** **Gal4**

```

TCATTGC-----TATATTGAAGTA CCGATTAGAAGCCGCCGAG CCGGCGACAGCCCTCCGA CCGGAAGACTCTCCTCCG TCGGTCC
TCATTGC-----AATATTGAAGTA CCGATCAGAAGCCGCCGAG CCGACGACAGCCCTCCGA CCGAATATTCCCTCCG TCGGTCC
TCAAG-----ATATTGAAATA CCGATGAGAAGCCGCCGAA CCGACGACAATTCCCGA CCGAACATTCTCCTCCG CGCGGCC
CCATTACTTCGCAATGTTGAAATA CCGATCAGAAGCTGCCGAC CCGATGACAGTACTCCGS CCGAAAAGTGTCTCCG TCGGAAG
**  ****  *****  *****  *****  *****  *****  *****  *****  *****  *****  *****  *****  *****  *****  *****

```

**Gal4**

```

CGCGTTCCTGAAACGCAGATGTGC CTCGCGCCGCACTGCTCCGAACAATAAGATTCTACAA-----TACTAGCTTTT--ATGG!
TGTGTCCCTTAA-CATCGATGTAC CTCGCGCCGCCCCTGCTCCGAACAATAAGATTCTACAAAGAA-TACTTGTTTTTTTATGG!
CGCGTCCCTGAA-CATAGGTACGG CTCGCGCCGCCCCTGCTCCGAACAATAAGATTCTACAAAGAAAGGTACTAATTTCT--ACGG!
CACGTCCCTGAT-TACTGAAGCGT CTCGCGCCGCCCATAACC CCGAACAATAAGATTCTACAAAGAAAGAA-TGCCTGTAGTG--GCAG!
**  ***  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *  *

```

**Mig1**

```

ATTGGCAGTAA----CCTGGCCCCACAAACCTT-CAAATTAACGAATCAAATTAACAACCATA-GGATGATAATGCGA-----!
ATAAGCAGCCC----ACTGACCCCATATACCTTTCAAACTATTGAATCAAATTGGCCAGCATA-TGGTAATAGTACAG-----!
ATAAACAGTCC----CCCGGCCCCACATACCTT-CAAATCGATGCGTAAACTGGCTAGCATA-GAATTTTGGTAGCAA-AATA!
ATGACAATTCTTGCCCT-CCCAATATACTTTGTTCCGTGTACAGCACACTGGATAGAACAATGATGGGTTGCGGTCAAGC!

```

Kellis *et al.* 2003 “Sequencing and comparison of yeast species to identify genes and regulatory elements”  
 Sequenced 4 closely related *Saccharomyces* genomes & identified conserved sequences in multiple alignments of orthologous sequences from the four species.

*Need species close enough to get reliable DNA alignment*

*Position of elements has to be conserved for detection*

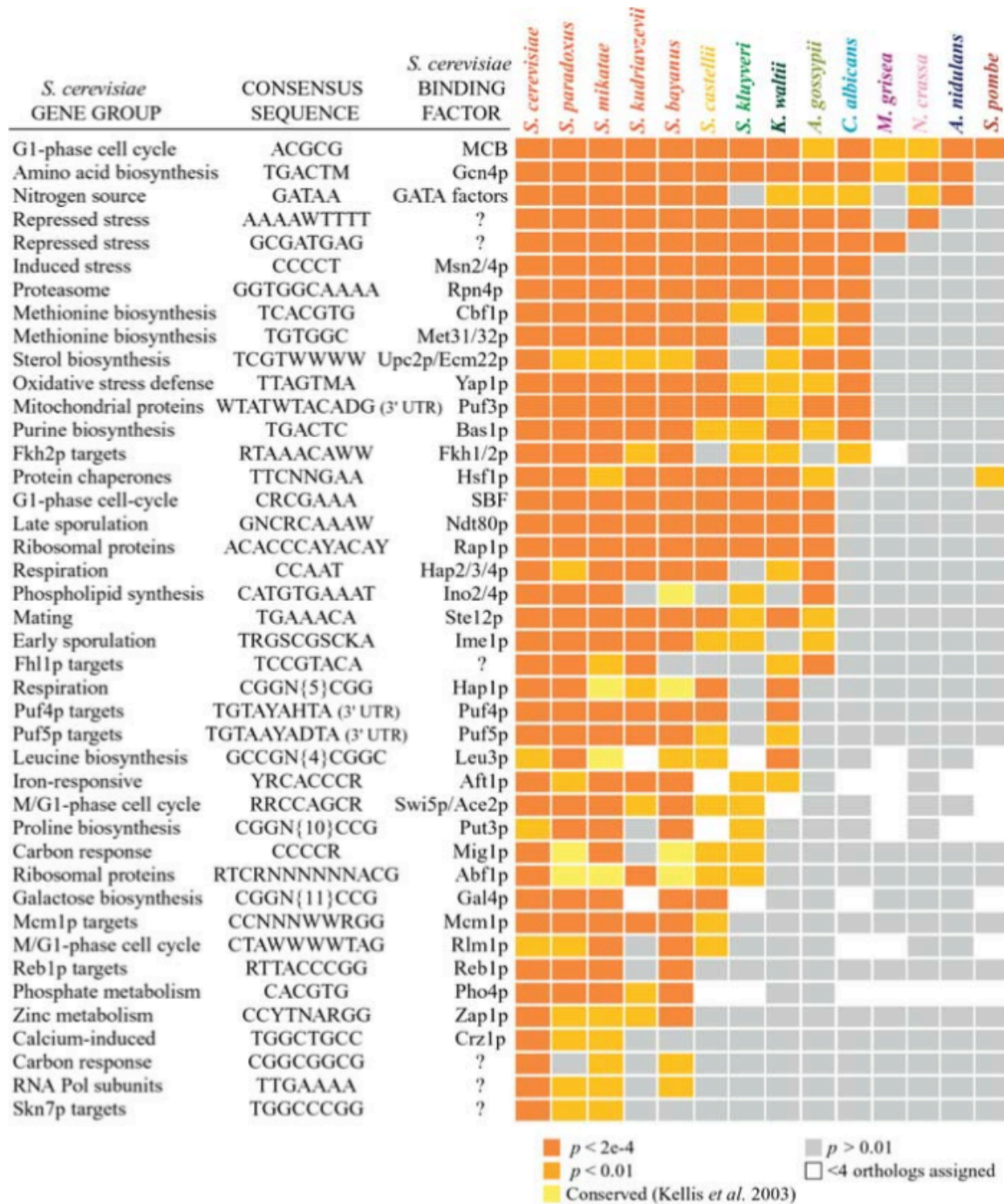
*(keep this in mind when we get to stabilizing selection at the end ...)*

## Identification of *cis*-regulatory elements

### Computational predictions:

1. Scan genome for matches to known matrix/consensus  
problem is that there are many nonfunctional in the genome - poor predictor of function
2. Phylogenetic footprinting: overly-conserved sequences in multiple alignments  
*Variation within element is typically lower than surrounding 'nonfunctional' DNA*
3. Network/module approach: Focus on groups of co-regulated genes to increase statistical power  
Look for statistically significant enrichment of sequences in  
the group of upstream regions from a group of co-regulated genes





## “Conservation and evolution of *cis*-regulatory systems in ascomycete fungi”

Gasch *et al.* 2004  
PLoS Biol

### Results:

- \* Many conserved elements are connected to similar gene groups over 100's of millions of years.

- \* Some gene groups show evidence of conserved co-regulation but evolved elements

- \* One example of co-evolved TF binding specificity and upstream sequence elements

## Identification of *cis*-regulatory elements

### Computational predictions:

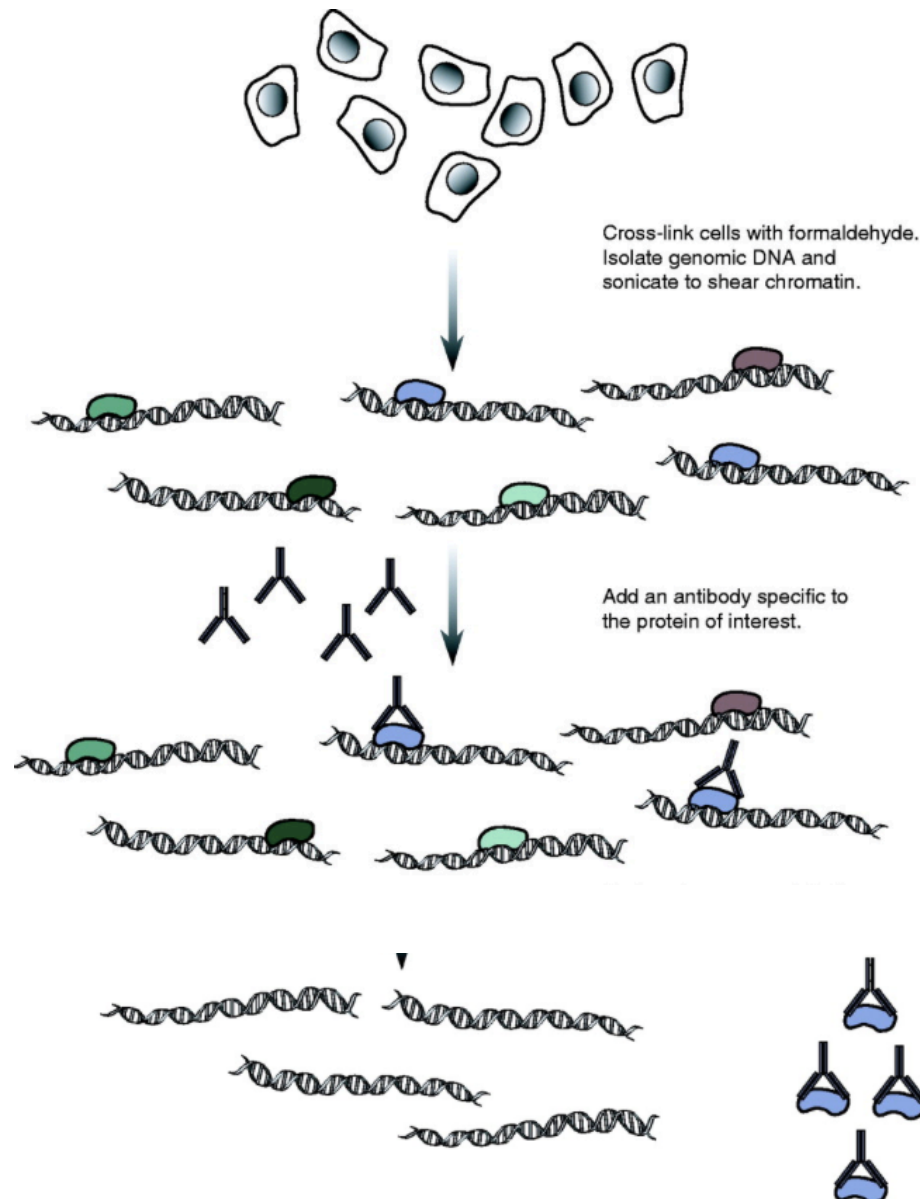
1. Scan genome for matches to known matrix/consensus  
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**Look for statistically significant enrichment of sequences in the group of upstream regions from a group of co-regulated genes**

### Experimental:

4. Chromatin immunoprecipitation (ChIP-chip or ChIP-seq) to identify binding loci genomewide  
*can do ChIP analysis across species or in one species then compare computationally*

# Chromatin-immunoprecipitation coupled to deep sequencing:

## ChIP-Seq:



1. Add crosslinker to cells
2. Lyse & shear DNA
3. IP protein of interest with antibody
4. Process recovered DNA & sequence

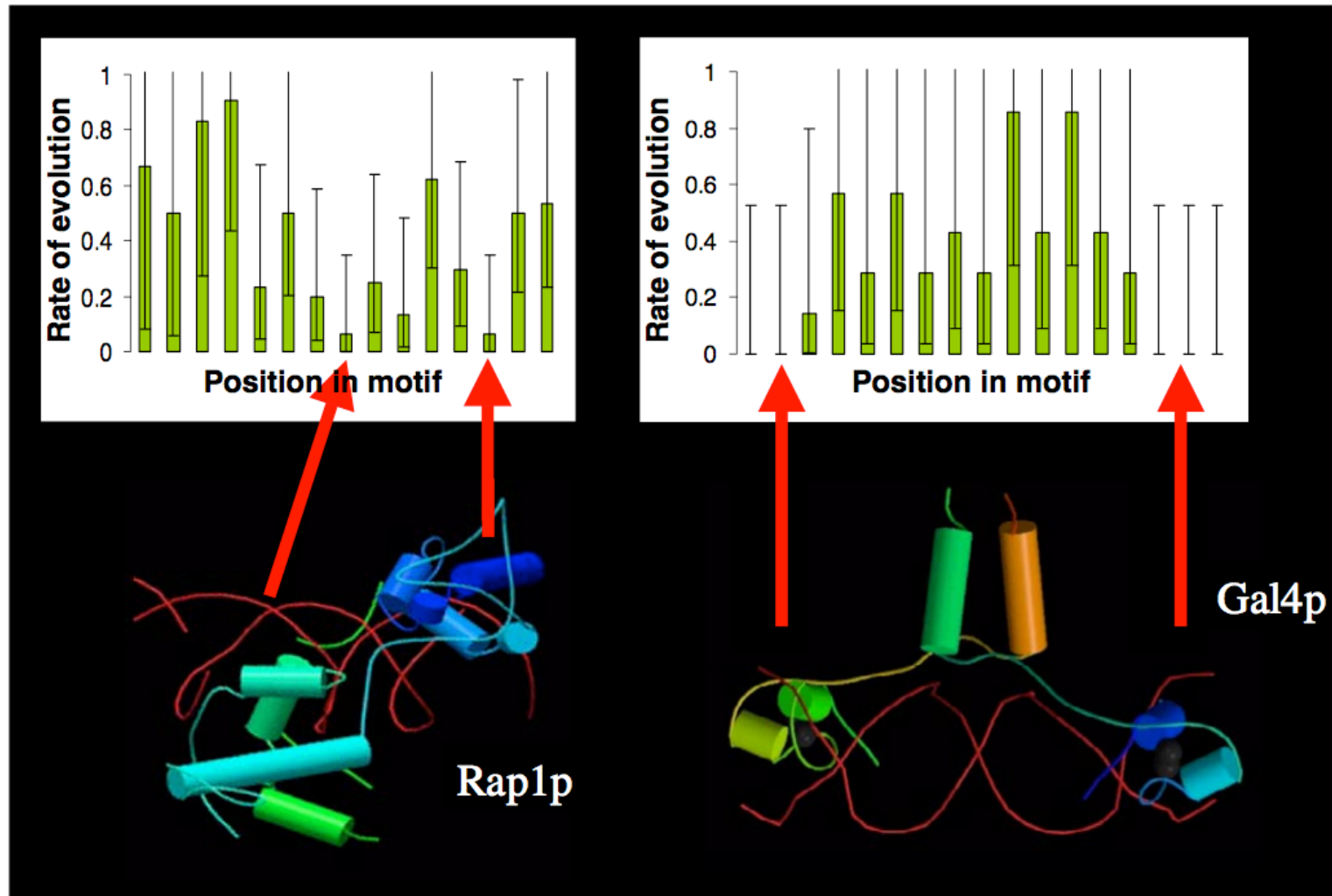
### *Lessons from ChIP*

- Best/most DNA recovery usually means highest TF-DNA affinity
- Often TFs bind DNA despite no recognizable ‘binding site’ in the region (note ChIP identifies a region bound, not a site)
- Many “low-occupancy” (e.g. weakly recovered) sites may be real binding that is non-functional

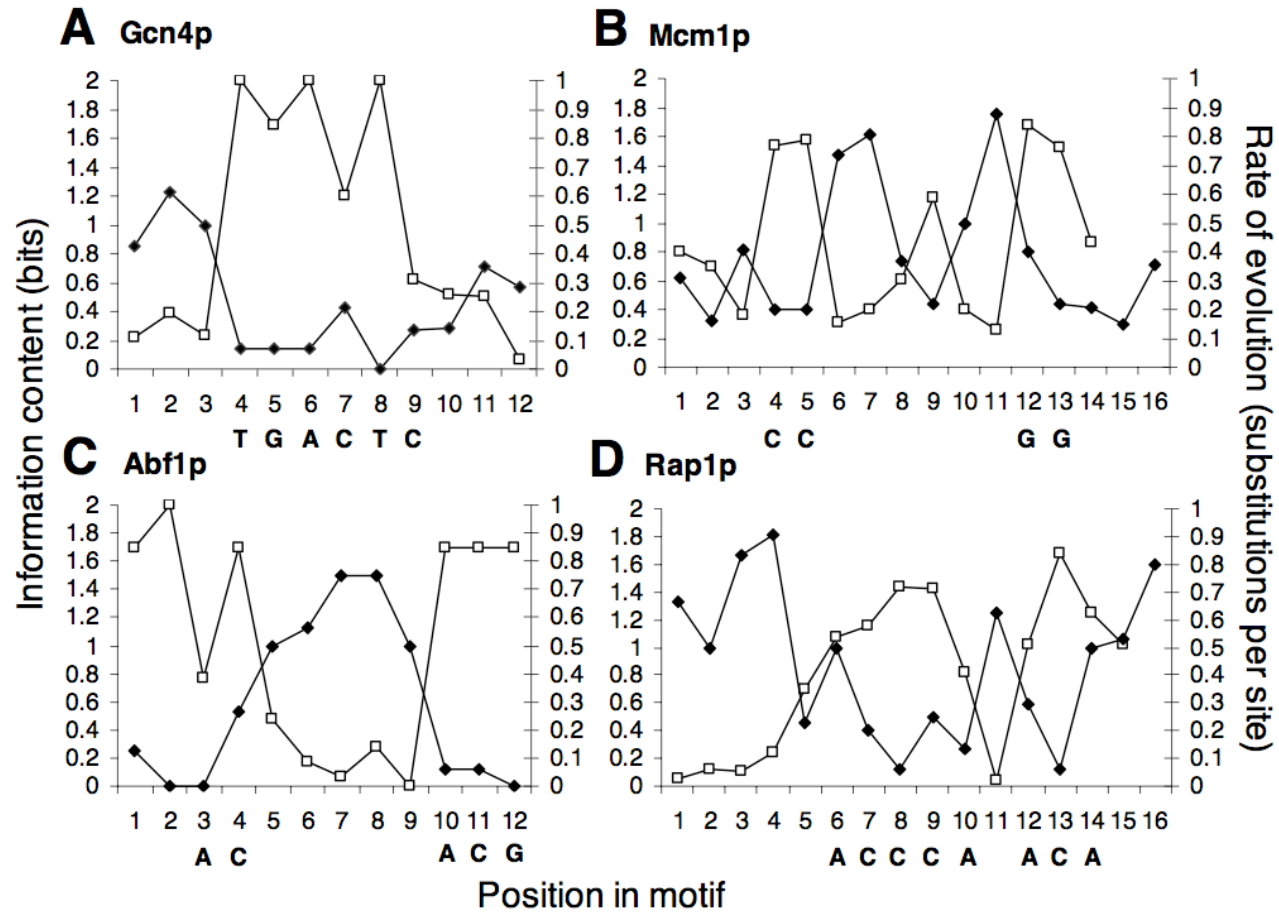
## What kinds of constraints act on TF binding sites?

1. Productive contact between protein-DNA (constraint on *sequence* of binding site)

Sites of contact evolve slower (under more constraint)



Variation within a site across species parallels variation across sites within a genome



Open symbols: Information content

Closed symbols: Substitutions per site

## What kinds of constraints act on TF binding sites?

1. Productive contact between protein-DNA (constraint on *sequence* of binding site)
2. Distance from transcription start site (constraint on *position* of the binding site)  
*also may be restricted by placement of nucleosome-depleted regions*

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## How do Regulatory Regions evolve?

1. Conserved regulation but evolution of regulatory regions (stabilizing selection)
  - Binding-site turnover: non-conserved sites but conserved regulation  
*Seems to be very prevalent across many organisms*

Ludwig *et al.* Nature. 2000



Four TFs act combinatorially  
To determine Eve2 patterns

Eve stripe 2 expression  
highly conserved across  
species.

None of 16 binding sites in  
stripe 2 enhancers  
is perfectly conserved  
across 13 species

**a**

```

TGCATAACAATGGAACCCGAACCGTAACTGGGACAGATCGAAAA..... mel
TGCATAACAATGGGCAAGGACCAGGGTTCGGTTCGCGAGATAAGGTTCTTTGACGGTTC pse
      *****

.....GCTGGCCTGGTTTCTCG....CTGTGTGTGCCGTGTTAATCCGTTTGCCA mel
CTTGACGGTTCCTTGAC.GGTTCCCTGTGTGCTCTGTCTGTGTTAATCCGTTTGCCA pse
      ***** ** **

-BC-4-      GT-3
TCAGCGAGATTATTAGTCAATTG.....CAGT.....TGCAG....CGTTTCGCTT mel
TCAGCAAGATTATTAGTCAATTTTCATATTTCCAGTCGAGTCGCAGTTTTGGTTTCACTT pse
*** * * ***** * * **

TC.....GTCCTCGTTTCACTT..... mel
TCTCCTTTGCCACTTCTTGCCTTGCTCATGTGGATGCCGATGCCGATGCCGTTGCCGT pse
*

.....TCGAGTTAGACTTTATTGCAGCATCTTGAACAATCGTCC mel
TGCCGTTGCCGTTGCCGACCGCAGGATTAGATTTTATTGCAGCATCTTGAACAATCAACT pse
*****

-GT-2-
CA.GTTTGGTAACACGCTG...TGCCA.....TACTTTC..... mel
GGAATTTGGTAACATGCTGCGCGGCCTAACCCTGGAGATTGCTCTACTTTCGCCTCAATT pse

.....ATTTAGACGGAAT.CGAGGGACCCTGGACTATAATCGCACAAACGAGACC..GG mel
GAATCGGAGTTAGCGGAAGACGGCGGACCCTTGCG.....ACCAAGG pse
* ** * * **

KR-4-
GTTGC....GAAGTCAGGGCATTCCGCCGATCTAGCCATCGCCATCTTCTGCGGGCGTT mel
GTTGTCTCTGGCCTCAGGAGTTTCCACAGTCAACGCTTTCGCT...GGTTTGTTTATT pse
***

-BC-2-
TGTTTGTGTTT...GCTGGGATTAGCCA.AGGGCTTGACTTGGAAATCCAATCCTGATCC mel
TGTTTGTGTTTTTAGCCAGGATTAGCCCCGAGGGCTTGACTTGGAAACCGA.CCAAAGCC pse
***** *****

C.....TAGCCCGATCCCAATCCCAATC mel
AAGGGCTTTAGGGCATGCTCAAGAGATCCCTATATCCCTATCCCTGTCGCGATCCCTAAA pse

CCAATCCC.TTGTCTT.TTTCATTAGAAAAGTCATAAAA.ACACATAATAATGA..TGTCG mel
CCGATCCCATTGGCAATTTTCATTAGAAAAGTCATAAAACACATAATAATGAGATGTCC pse
***** * * * * **

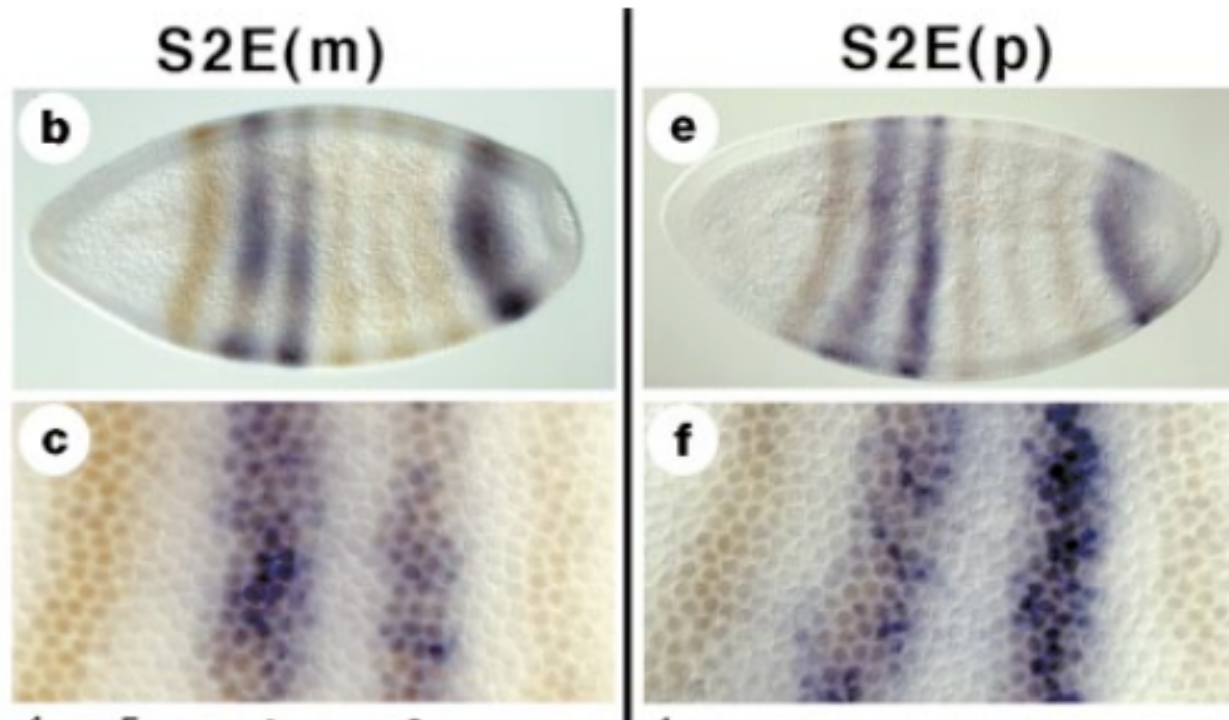
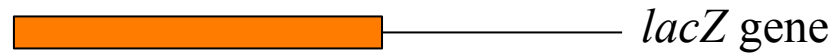
-BC-1-      HB-3      -BC-1-
-BC-1-      KR-3-
AAGGGATTAGG....GGCGCGCAGGTCAGGC...AACGCAATTAACGGACTA 504 mel
AAGGGATTAAGATTAAGGGACGCACACACAGGCAGCAGGATCATTAAACGGACTA 691 pse
***** ** * * * *

```

Ludwig *et al.* Nature. 2000

**Evidence for stabilizing selection in a eukaryotic enhancer element.**

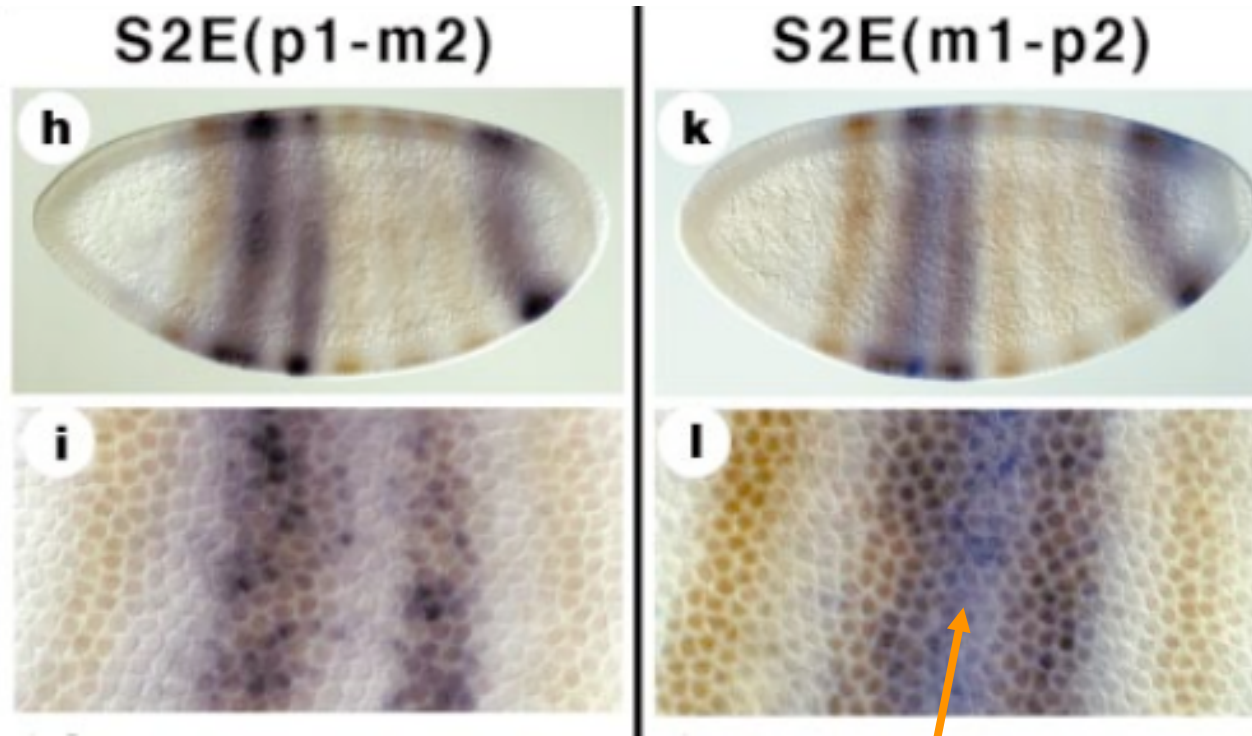
Native *D. pseudoobscura* enhancer works well in *D. melanogaster*



Ludwig *et al.* Nature. 2000

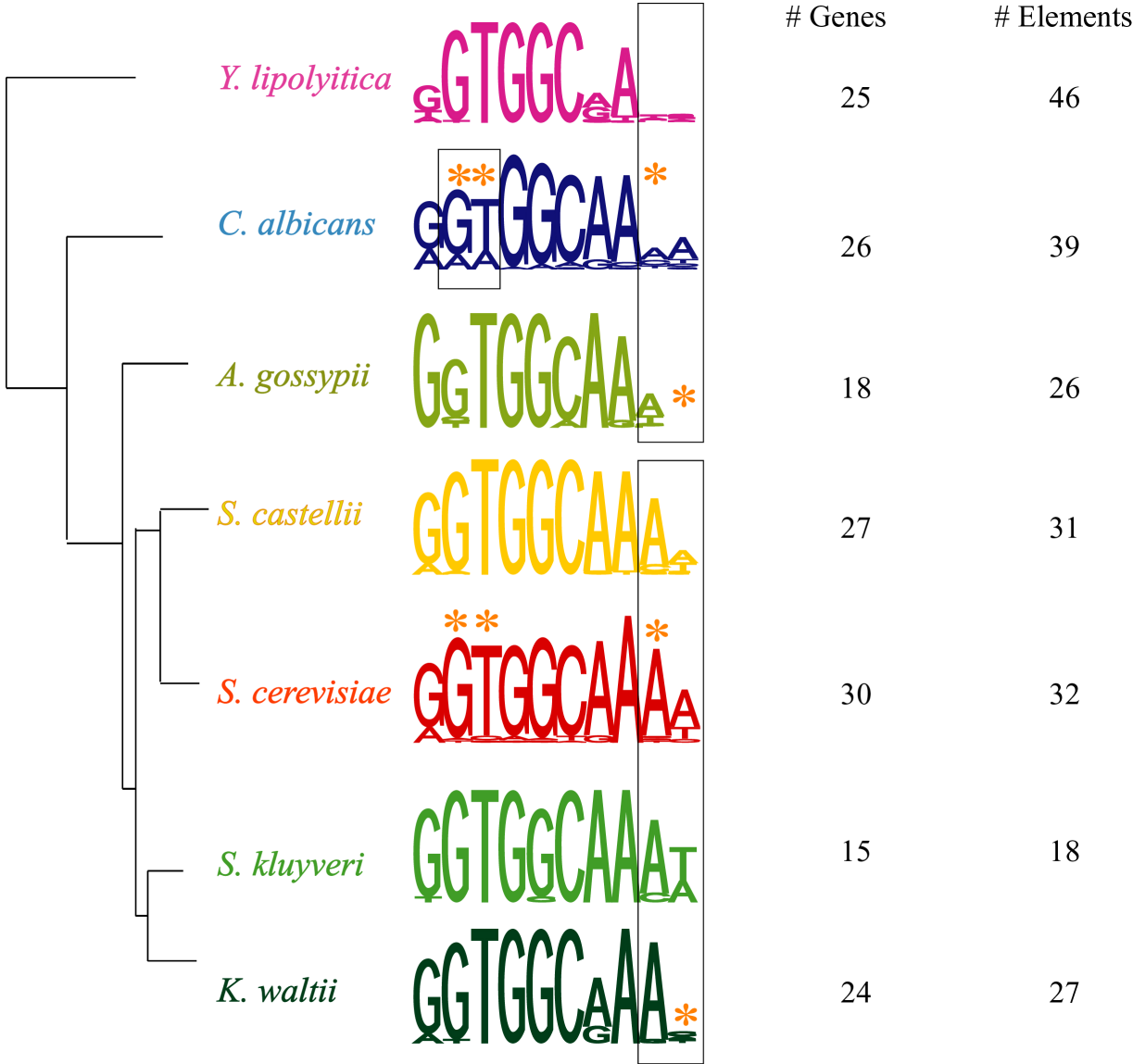
**Evidence for stabilizing selection in a eukaryotic enhancer element.**

But hybrid enhancers (mel-pseudo or pseudo-mel from 5' to 3') are defective



They argue for stabilizing selection and binding-site turnover across the enhancer 20

# Co-evolution of Rpn4 sites upstream proteasome genes & Rpn4 binding specificity



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2. Distance from transcription start site (constraint on *position* of the binding site)  
*also may be restricted by placement of nucleosome-depleted regions*
3. Spacing between elements if cooperative TF interactions (constraint of *position*)

## How do Regulatory Regions evolve?

1. Conserved regulation but evolution of regulatory regions (stabilizing selection)
  - Binding-site turnover: non-conserved sites but conserved regulation  
*Seems to be very prevalent across many organisms*
  - Co-evolution between binding site and TF specificity