

BEAST: Bayesian Evolutionary Analysis by Sampling Trees

A STEP-BY-STEP TUTORIAL FOR PHYLOGEOGRAPHIC INFERENCE IN CONTINUOUS SPACE

This step-by-step tutorial guides you through a continuous space phylogeographic analysis for reconstructing the spatial dynamics of a large-scale rabies virus outbreak among North American raccoons. The data set consists of 47 N gene sequences of raccoon rabies virus (RABV), isolated at different time points (1982-2004) throughout the USA [Biek *et al.*, 2007]. Here we describe a full Bayesian framework for phylogeographic reconstruction developed in [Lemey *et al.*, 2009].

To undertake this tutorial, you will need to download the following software packages in a format compatible with your operating system (available for Linux/UNIX, Mac OS X and Windows):

- **BEAST** - this package contains the BEAST program, BEAUti and a couple of utility programs. At the time of writing, the current version is v1.8.0. It is available for download from <http://beast.bio.ed.ac.uk>
- **Tracer** - this program is used to explore the output of **BEAST** (and other Bayesian MCMC programs). It graphically and quantitatively summarizes the empirical distributions of continuous parameters and provides diagnostic information. At the time of writing, the current version is v1.6. It is available for download from <http://tree.bio.ed.ac.uk/software/tracer/>
- **FigTree** - this is an application for displaying and printing molecular phylogenies, in particular those obtained using **BEAST**. At the time of writing, the current version is v1.4.2. It is available for download from <http://tree.bio.ed.ac.uk/software/figtree/>
- **SPREAD** - this is an application for the visualizing and quantifying support for phylogeographic analyses performed with **BEAST**. At the time of writing, the current version is v1.0.6. It is available for download from <http://www.kuleuven.be/aidslab/phylogeography/home.html>
- **Google Earth** - virtual globe software that can be used to visualize the KML output from SPREAD in an interactive fashion. It is available for download from <http://www.google.com/earth/>
- **BEAGLE library** - high-performance *library* that can perform the core calculations at the heart of most Bayesian and Maximum Likelihood phylogenetics packages. It is available for download from <https://code.google.com/p/beagle-lib/>

Suggested text editors for editing xml files:

- **Geany** (Linux/UNIX):
<http://www.geany.org/Download/Releases>
 - **TextWrangler** (Mac OS X):
<http://www.barebones.com/products/textwrangler/download.html>
 - **Notepad++** (Windows):
<http://notepad-plus-plus.org/download/v6.6.8.html>
-

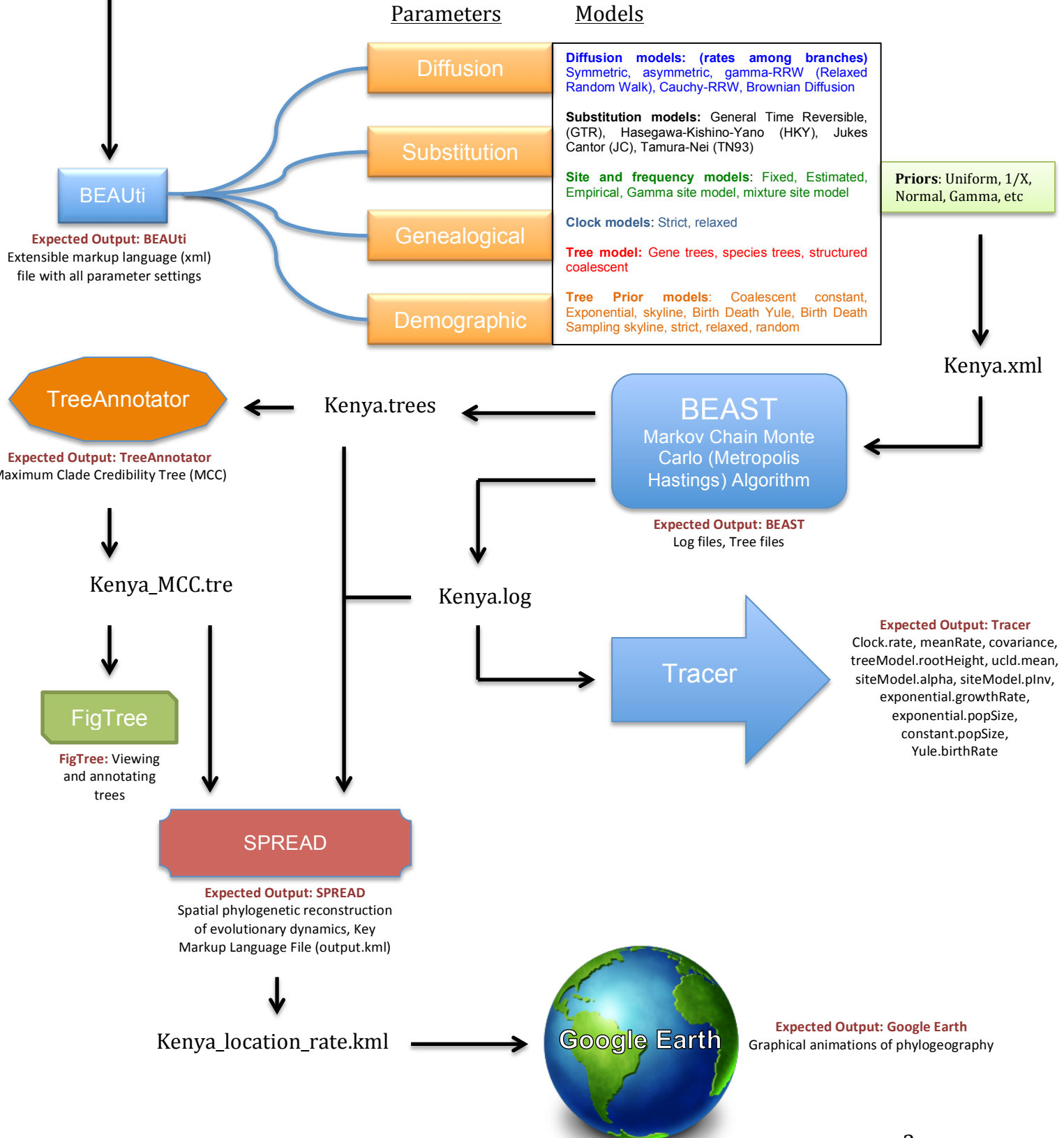
ANALYSIS PIPELINE

Input: FASTA/ NEXUS file (e.g. Kenya.fasta)

EX000001_Nairobi_2003
EX000002_Mombasa_2004
EX000003_Kisumu_2005
EX000004_Nakuru_2006

ATCATGGGGCCTGAAA
A-CAT---GGCCCTGAAT
ATCATGCGCGATGAAA
ATACTCGCGCTTCTAT

	Accession number
	Sampling location
	Collection date

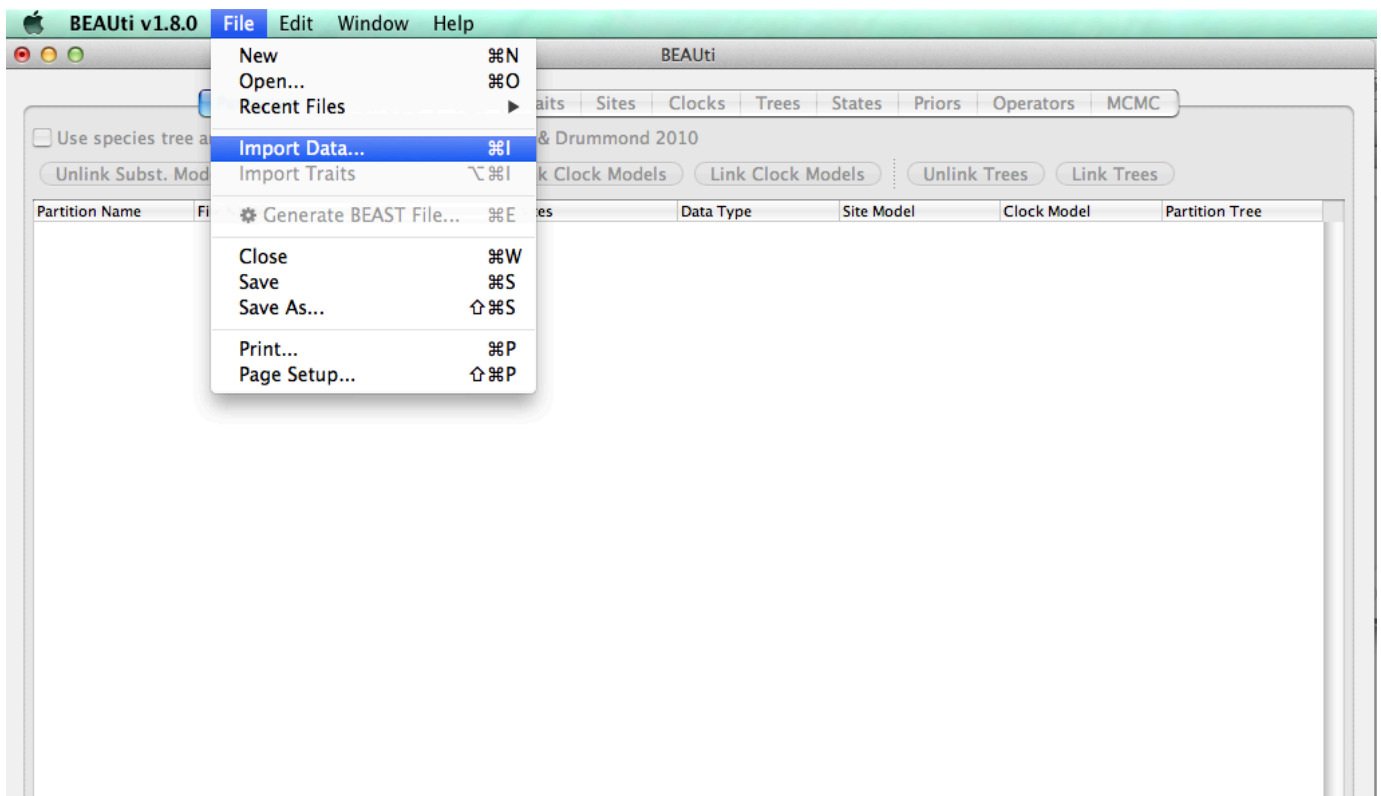


RUNNING BEAUti

BEAUti is a user-friendly utility for setting model parameters for Bayesian analysis in BEAST.

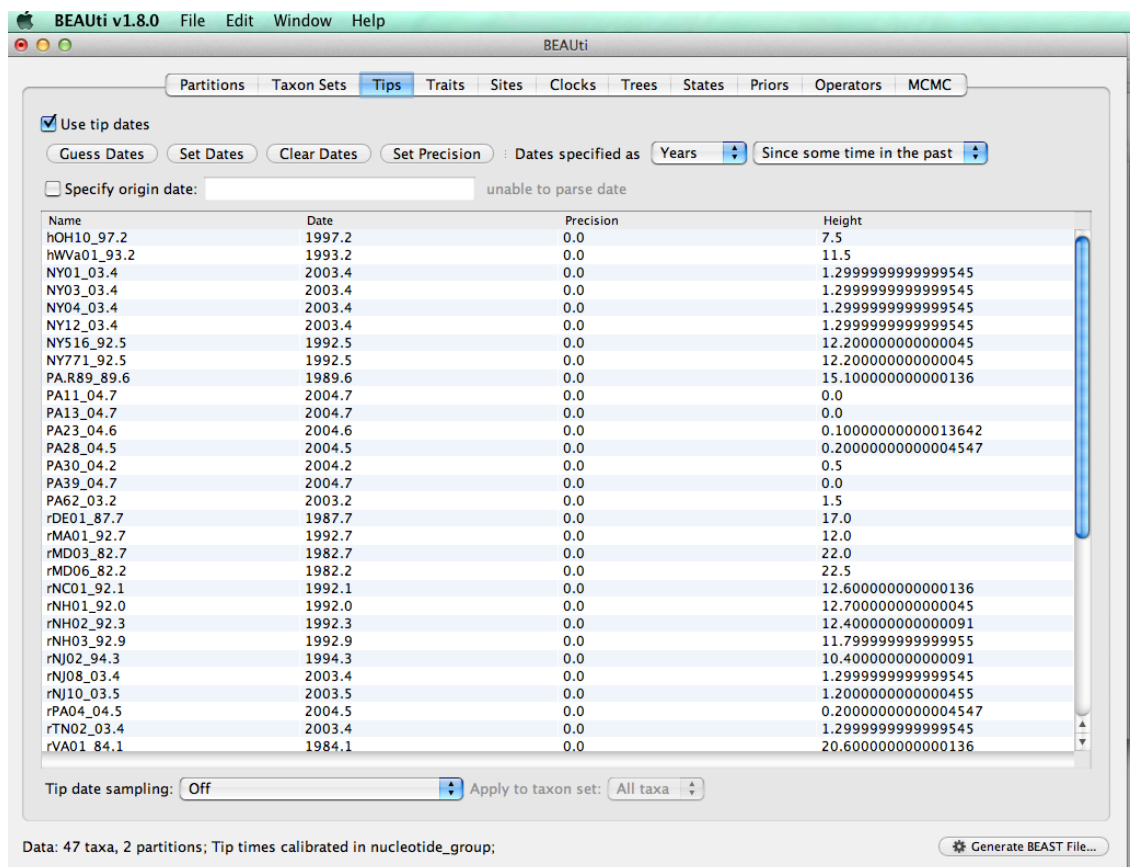
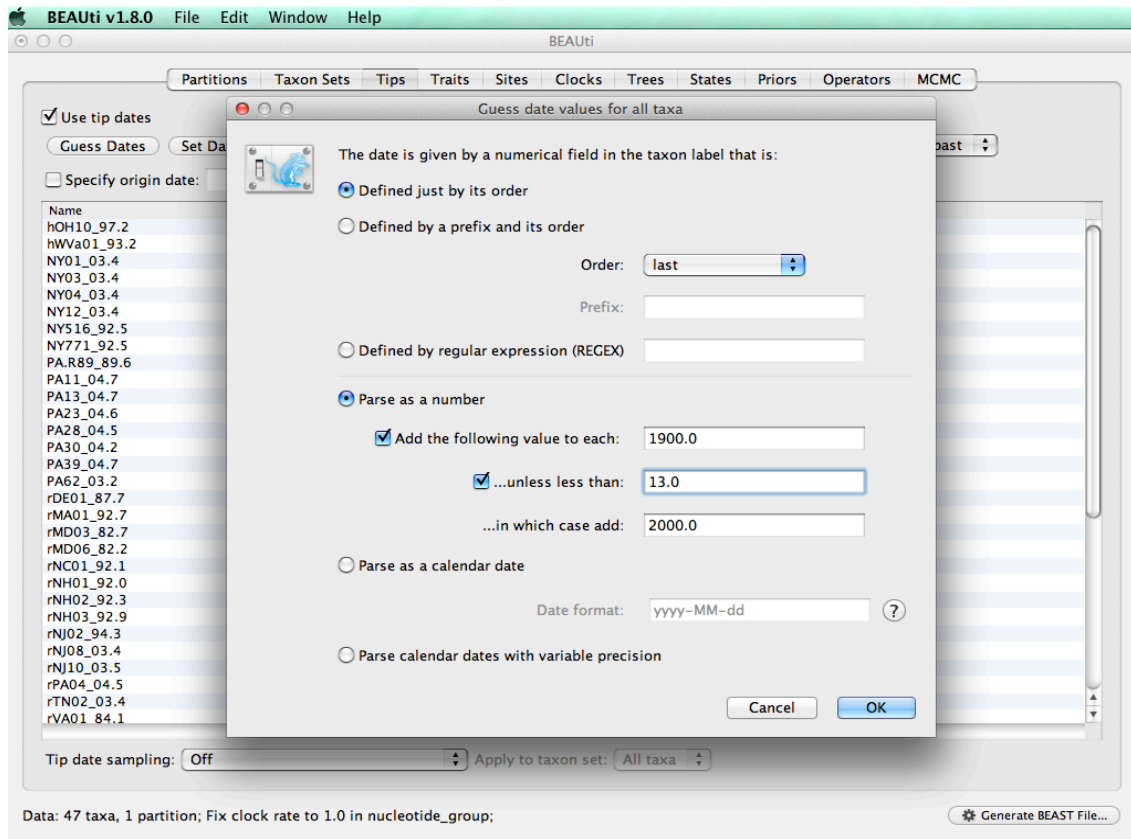
1. Partitions Tab:

- Load the **RacRABV.fasta** file by selecting **Import Data** from the **File** menu. Once loaded, the sequence data will be listed under **Partition Name**.



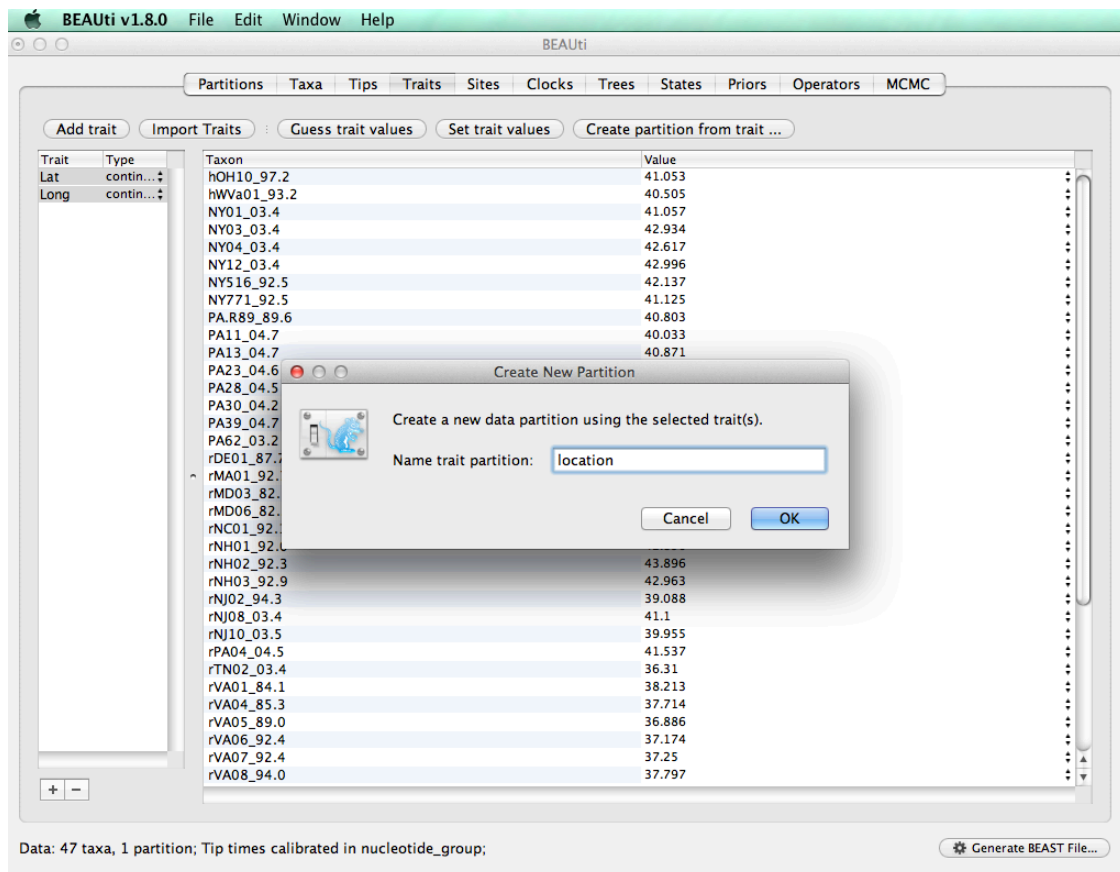
2. Tips Tab:

- Select '**Use tip dates**' option. Click on '**Guess Dates**'. In the dialogue box that appears keep the default '**Defined just by its order**' and set the '**Order**': to **last**.
- Go to '**Parse as a number**' and select '**Add the following value to each:**'. Keep the default **1900.0**. Select '**unless less than:**'. Keep the default **15.0** and '**...in which case add:**' **2000.0**.



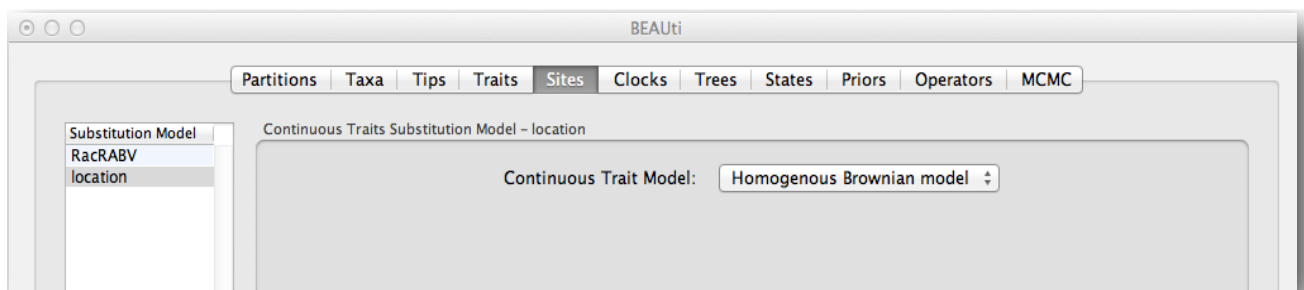
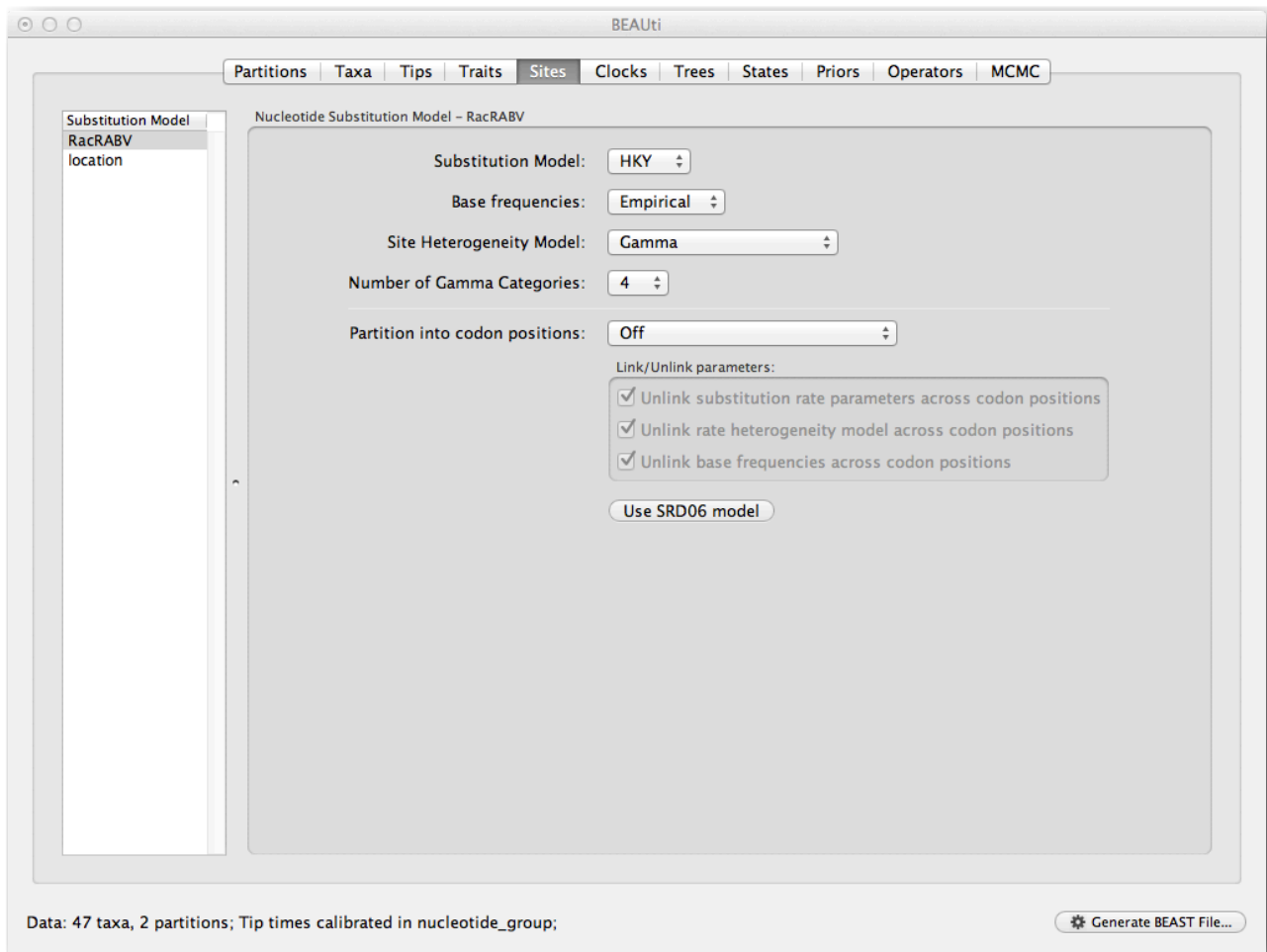
3. Traits Tab:

- Click '**Add trait**'. In the dialogue box that appears, select '**Import trait(s) from a mapping file format**' and click **OK**.
- Load the **LatLong.txt** tab-delimited file, which contains latitudes and longitudes associated with each sequence. Click **OK**.
- Select both the **Lat** and **Long** traits and click the '**Create partition from trait...**' button. Enter the name **location** for this partition and click **OK**. This new partition will now appear in the Partitions tab as a continuous Data Type.



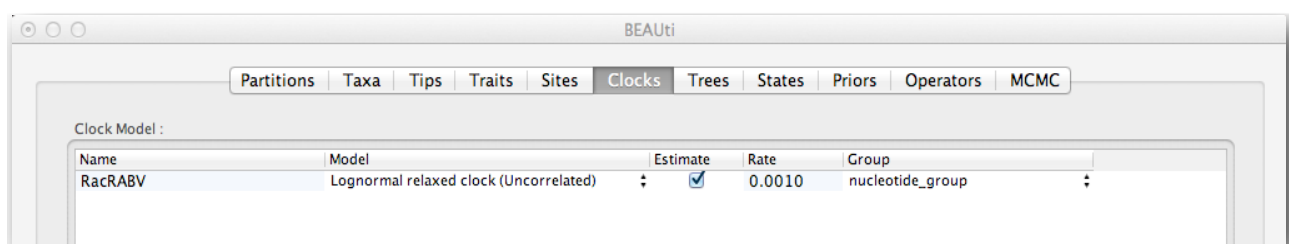
4. Sites Tab:

- Keep **Substitution Model** as the default **HKY**
- Set **Base frequencies** to **Empirical**, and
- Set **Site Heterogeneity Model** to **Gamma** distributed rate variation among sites.
- Keep the **Number of Gamma Categories** at **4**.
- Click on **location** in the **Substitution Model** window on the left and keep the default **Homogenous Brownian model**.



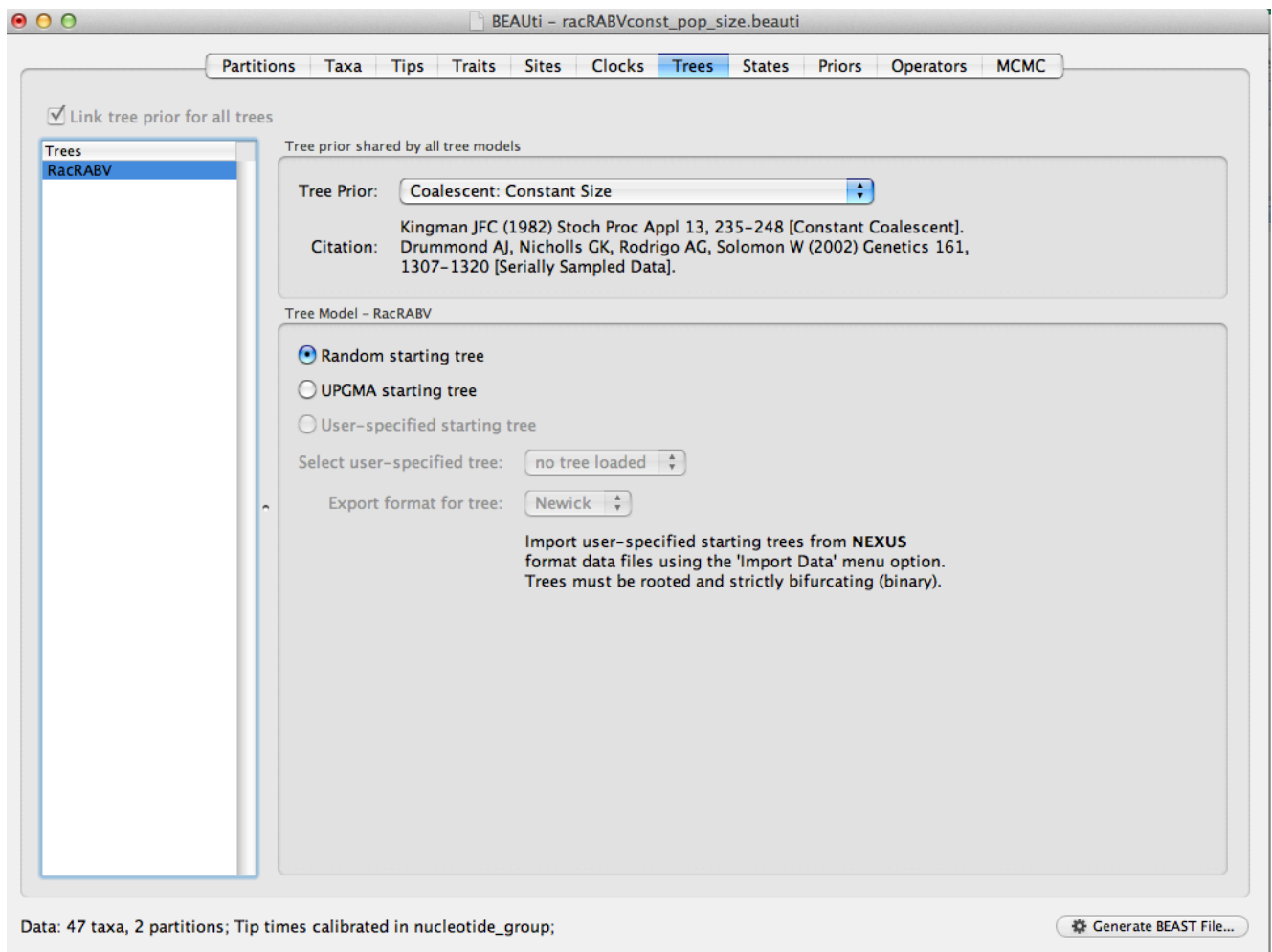
5. Clocks Tab:

- Select the model **Lognormal relaxed molecular clock (Uncorrelated)** and set the initial **Rate** value to **0.0010**.



6. Trees Tab:

- On the **Tree Prior**, select the **Coalescent: Constant Size** model and keep the other default settings.

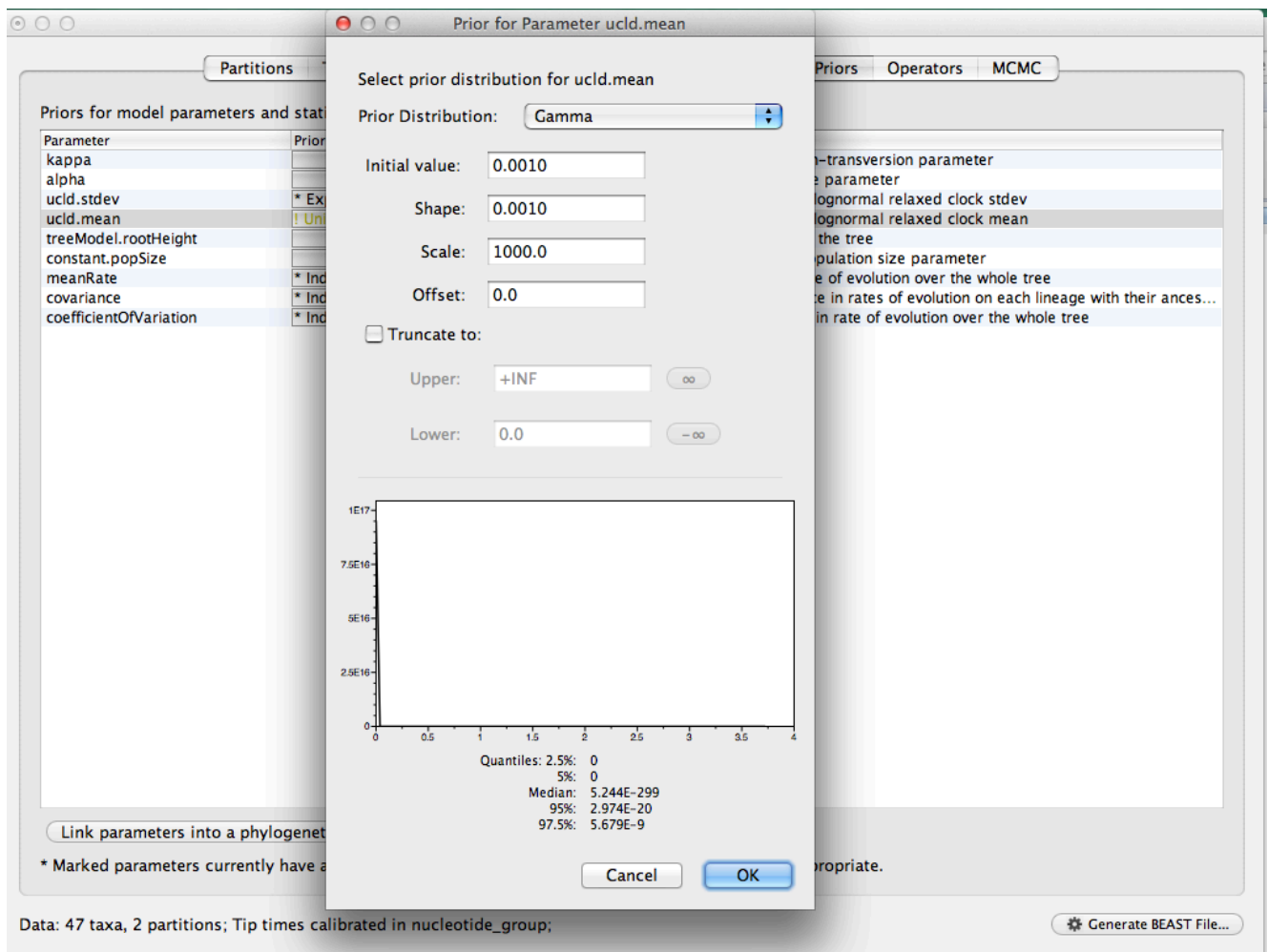


7. States Tab:

- Click on the **location** partition on the left, and select the option '**Reconstruct states at all ancestors**'.

8. Priors Tab:

- Priors that have not been set yet appear in red (e.g. **uclid.mean**).
- Click on the prior for the **uclid.mean** parameter.
- In the dialog window that appears, set the '**Prior Distribution**' to **Gamma** with **initial value = 0.001**, **shape = 0.001** and **scale = 1000**. Click **OK**.



9. MCMC Tab:

- Set the **Length of Chain** to **1 000 000**
- Set both sampling frequencies (**Echo state to screen every:** and **Log parameters every:**) to **10 000**.
- Set **File name stem** to **RacRABV_homogeneous**.
- Select **Create operator analysis file**.
- At the bottom, select **Perform marginal likelihood estimation (MLE) using path sampling/ stepping-stone sampling**. Keep default settings.
- Click the **Generate BEAST File** at the bottom of the page.
- Click **Continue**. Choose the name of the file and the directory where it will be saved. Click **Save**.

Leave the BEAUti window open, so that you can change the values and re-generate the BEAST file if necessary.

Open the **RacRABV_homogeneous.xml** file in an xml text editor to manually add an additional diffusion model parameter (**treeDispersionStatistic**), which *cannot* be specified by this version of **BEAUti**. This parameter records the *rate of diffusion* along each branch, and provides a diffusion estimate in km/yr.

Add the following block (e.g., by copy- pasting) just before the operators block in the xml:

```
<treeDispersionStatistic id="dispersionRate" greatCircleDistance="true">
    <treeModel idref="treeModel"/>
    <multivariateTraitLikelihood idref="location.traitLikelihood"/>
</treeDispersionStatistic>

<!-- END Multivariate diffusion model -->

<!-- Define operators -->
```

And also add a reference to it in the log file:

```
<log id="fileLog" logEvery="100" fileName="RacRABV_homogeneous.log"
overwrite="false">
```

```
<posterior idref="posterior"/>
```

```
<prior idref="prior"/>
```

....

```
<treeDispersionStatistic idref="dispersionRate"/>
```

....

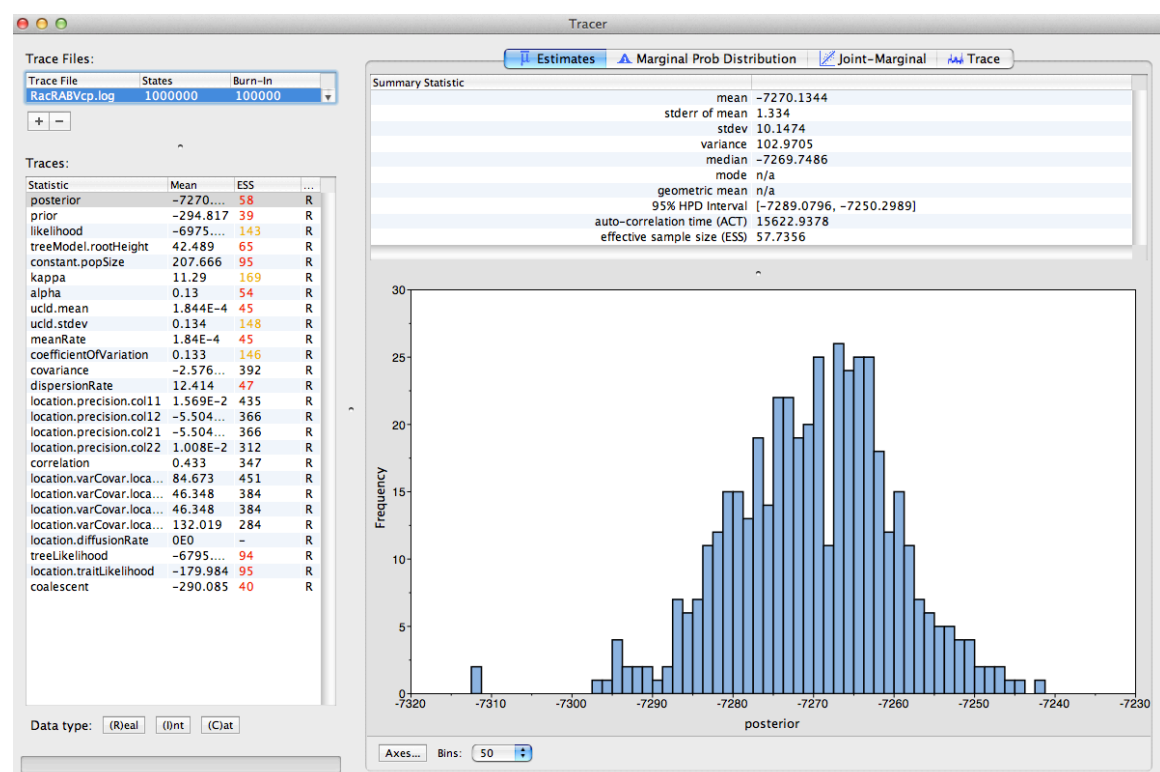
Save the file.

RUNNING BEAST

- Click on **Choose File...** and open **RacRABV_homogeneous.xml**.
- Select the '**Use BEAGLE library if available**' option.
- Keep default settings and select **Run**.

ANALYSING THE BEAST OUTPUT

- Execute **Tracer**
- Select **Import Trace File...** from the **File** menu and load the log file created by BEAST (**RacRABV_homogeneous.log**).



Answer the following questions

- a) At what rate did RABV spread in the raccoon population in North America? (Note that the dispersionRate statistic is measured in km/year).
- b) What is the age of most recent common ancestor (TMRCA)?
- c) What does it mean when the EES values are displayed, in red, yellow or black? How can we improve ESS estimates?

SUMMARISING THE TREES

- Open **TreeAnnotator**. Click on **Choose File...** next to **Input Tree File** and open **RacRABV_homogeneous.trees**
- **Burn-in** - This is the number of trees in the input file that should be excluded from the summarization. This value is given as the number of trees rather than the number of steps in the MCMC chain. Thus for a chain length of 10 000 000 steps, sampling every 10 000 steps will result in 1000 trees. To obtain a 10% **Burnin**, set the value to **100**.
- **Posterior probability limit** - This is the minimum acceptable posterior probability for a node. Keep the **default** is **0.0** so all nodes will be annotated.
- **Target tree type** – Keep the default **Maximum clade credibility tree** option. TreeAnnotator will examine every tree in the Input Tree File and select the tree that has the highest sum of the posterior probabilities of all its nodes.
- **Node heights** - Keep the default **Median heights**.
- **Output File** – Click on **Choose File...** next to **Output File**. Enter a name for the output file (e.g. **RacRABV_homogeneous_mcc.tre**).
- Click **Run**

VIEWING THE ANNOTATED TREE

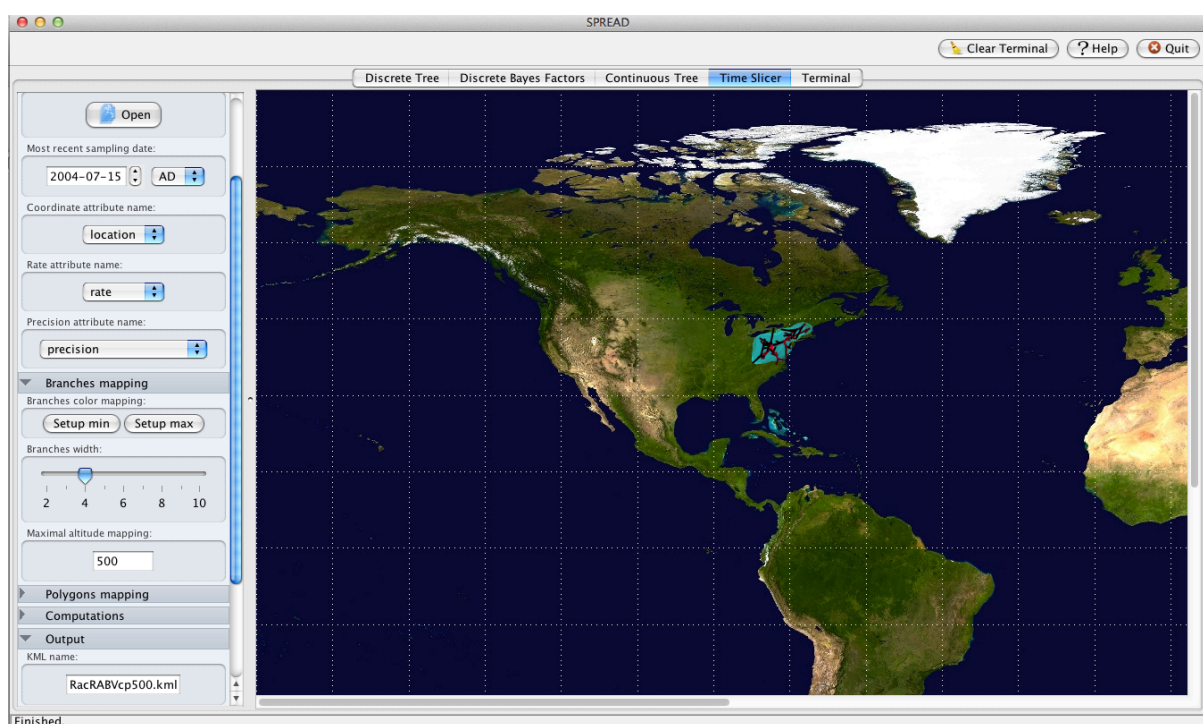
- Open **Figtree**. Under the **File** menu, select **Open**.
- Select and open the **RacRABV_homogeneous_mcc.tre** file created by TreeAnnotator
- Check next to and open the **Branch Labels tab** on the left.
- Next to **Display**, select **Posterior** from the dropdown menu.
- We now want to display bars on the tree to represent the estimated uncertainty in the inferred date for each node. Check and open the **Node Bars tab** and choose **Display: height_95%_HPD** from the dropdown menu.

- To display a time scale for this phylogeny, deselect the **Scale bar tab**.
- For appropriate scaling, open the **Time Scale tab**, and set the **Offset by:** to **2004.7**, the **Scale Factor:** to **-1.0**.
- Tick and open the **Scale Axis tab**. Check the **Reverse axis** option.
- In the **Layout tab**, tick **Align Tip Labels**.
- Finally, open the **Appearance tab** and set the **Line Weight** to **3**.

None of the options actually alter the tree's topology or branch lengths in anyway, so feel free to explore the options and settings. You can also save the tree with all your settings so that you can reload it into FigTree.

EVALUATING DIFFUSION RATE VARIATION

- Open **SPREAD**
- Click on the **Time Slicer tab**.
- Under **Load slice heights/ tree file**, open **RacRABV_homogeneous_mcc.tre**.
- Next to **Load trees file**, open **RacRABV_homogeneous.trees**
- Set the **Most recent sampling date** to **2004-07-15**.
- Under **Coordinate attribute name**, select **location**.
- Under **Rate attribute name**, keep the default **rate**.
- Under the **Branches mapping tab**, set the **Maximal altitude mapping** to **500**.
- Under the **Output tab**, give an appropriate **KML name** (e.g. **RacRABVcp500.kml**).
- Click **Plot** and **Generate**.



Biek R, Caroline Hendorson J, Waller LA, Rupprecht CE, Real LA (2007). **A high-resolution genetic signature of demographic and spatial expansion in epizootic rabies virus.** Proceedings of the National Academy of Sciences, USA 104(19): 7993–7998.

Lemey P, Rambaut A, Drummond AJ, Suchard MA (2009). **Bayesian Phylogeography Finds Its Roots.** PLoS Computational Biology 5(9): e1000520. doi:10.1371/journal.pcbi.1000520

Recommended Literature

BEAST2 manual

<http://beast2.org/>

Selecting proper priors

<http://code.google.com/p/beast-mcmc/wiki/ParameterPriors>

Lemey P *et al.*, 2010. Phylogeography Takes a Relaxed Random Walk in Continuous Space and Time.

<http://mbe.oxfordjournals.org/content/27/8/1877.full>

BEAST: Bayesian evolutionary analysis by sampling trees. Alexei J Drummond and Andrew Rambaut. 2007.

<http://www.biomedcentral.com/1471-2148/7/214>

SPREAD Tutorial.

http://www.kuleuven.be/aidslab/phylogeography/tutorial/spread_tutorial.html

Phylogeographic Inference in continuous space: A hands-on Practical.

Suchard M and Lemey P. July 2013.

https://perswww.kuleuven.be/~u0036765/SISMID/handouts_files/2013_SISMID_12_16.pdf